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Global Biogeochemical Cycles

Supporting Information for

A Mass Budget for Mercury and Methylmercury in the Arctic Ocean

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Introduction

The Supporting Information contains two texts, nine tables, and four figures.

Rate constants

		median (d ⁻¹)	, methylation and	$\frac{1}{1}$ intering define the form	Defenses of
	avg±stav (a)	median (d)	min-max (a)	avg±stuv (m E)	кенегепсе
Methylation					
Unfiltered (dark)	0.0068±0.0039*	0.0050*	0.0006 - 0.013*		[Lehnherr et al., 2011]
Unfiltered (light)	0.0179 ± 0.0197	0.0085	< 0.0002 - 0.060		[Monperrus et al., 2007]
Unfiltered (dark)	0.0154 ± 0.0119	0.0130	< 0.0002 - 0.038		[Monperrus et al., 2007]
Filtered (light)	0.0072 ± 0.0072	0.0040	< 0.0002 - 0.017		[Monperrus et al., 2007]
Filtered (dark)	0.0056 ± 0.0046	0.0060	< 0.0002 - 0.013		[Monperrus et al., 2007]
Unfiltered (dark)			< 0.0001-0.001		[<i>Sharif et al.</i> , 2014]
Unfiltered (light)			< 0.0001-0.004		[<i>Sharif et al.</i> , 2014]
Filtered (light)			< 0.0001-0.003		[Sharif et al., 2014]
Filtered (dark)			< 0.0001		[Sharif et al., 2014]
Unfiltered (dark)	0.0025		Max 0.004		[Schartup et al. 2015]
n/a	0.13				[Mason and Sullivan, 1999]
Demethylation					
Unfiltered (dark)	0.36±0.09*		0.23 - 0.59*		[Lehnherr et al., 2011]
Filtered (light)	$0.04^{a} *$			1×10 ⁻³ *	[Lehnherr et al., 2011]
Filtered (light)	$0.2^{b} *$				[Lehnherr et al., 2011]
n/a			< 0.09 - 0.43		[Whalin et al., 2007]
Unfiltered (light)	0.16±0.05	0.159	0.048 - 0.245		[Monperrus et al., 2007]
Unfiltered (dark)	0.06 ± 0.03	0.059	< 0.015 - 0.109		[Monperrus et al., 2007]
Filtered (light)	0.09 ± 0.04	0.090	0.033 - 0.139		[Monperrus et al., 2007]
Filtered (dark)	0.05 ± 0.03	0.064	< 0.015 - 0.088		[Monperrus et al., 2007]
Unfiltered (light)				$3.2(\pm 1.0) \times 10^{-3}$	[Black et al., 2012]
Unfiltered (light)				$9.9(\pm 26.0) \times 10^{-3}$	[Black et al., 2012]
Unfiltered (dark)			0.07-0.21	× ,	[Sharif et al., 2014]
Unfiltered (light)			0.06-0.55		[Sharif et al., 2014]
Filtered (light)			0.08-0.29		[Sharif et al., 2014]
Filtered (dark)			< 0.02-0.13		[Sharif et al., 2014]
Dimethylation					
Unfiltered (dark)			nd-0.0016*		[Lehnherr et al., 2011]
Me ₂ Hg decomposition					
Light				<1×10 ⁻³	[<i>Black et al.</i> , 2009]
n/a			0.0002		[<i>Mason et al.</i> , 1995]
Light			0.32 - 1.64		Mason and Sullivan, 1999]
Dark			0.16 - 0.22		[Mason and Sullivan, 1999]

Table S1. Compilation of measured rate constants for Hg^{II} methylation and MeHg demethylation in seawater.

*Arctic data, ^a 9 days incubation, ^b 24 hours incubation

Budgets

A) Solids budget



B) Hydrologic budget



Figure S1. A: Solids budget for the Arctic Ocean in Tg a⁻¹. Red arrows denote external sources, green arrows denote internal sources, black arrows denote internal fluxes and blue arrows represent losses. The fraction of solids from rivers and NPP remaining after remineralization at each depth is 50% in the PML, 30% in the subsurface water and 1% in the deep ocean [*Cai et al.*, 2010; *Moran et al.*, 1997; *Rachold et al.*, 2004]. Solids from erosion are assumed not to remineralize. B: Hydrologic budget for external input and losses and internal circulation in the Arctic Ocean (×10¹² m³ a⁻¹).



Figure S2. Empirical mass budget for the Arctic Ocean for inorganic Hg species including uncertainty ranges for masses and fluxes not given in Table S2. Red arrows are external sources, black arrows are internal fluxes and blue arrows are losses. Punctuated lines describe net fluxes. Masses are in Mg and fluxes in Mg a⁻¹.



Figure S3. Empirical mass budget for the Arctic Ocean for organic Hg species including uncertainty ranges for masses and fluxes not given in Table S2. Red arrows are external sources, black arrows are internal fluxes and blue arrows are losses. The punctuated lines describe the net flux of biotic methylation/demethylation. Masses are in Mg and fluxes in Mg a⁻¹.

Observations

Text S1. Information on North Atlantic inflow data

Total inorganic Hg and methylated Hg concentrations from the CLIVAR A16N Cruise Track in the North Atlantic for stations occupied in August 2013. Measurements and analytical details were the same as Sunderland et al. (2009). The same depth intervals as in the Arctic were used to calculate average depth concentrations. Data are from station 007 (61.6 N; 20.0 W) and station 015 (58.0 N; 20.0W)(n=24).

0-20 meter, surface:	0.48±0.08 pM Hg,	42±25 fM methylated Hg
20-200 meter, subsurface:	0.50±0.06 pM Hg,	38±18 fM methylated Hg
>200 meter, deep ocean:	0.6±0.06 pM Hg,	153±99 fM methylated Hg

Text S2. Canadian Arctic Archipelago Seawater Samples Collected by I. Lehnherr et al. Seawater samples were collected at 17 stations across the Canadian Arctic Archipelago (Figure 1), in 2006, 2007 and 2010, between 7-August and 16-October while aboard the CCGS Amundsen icebreaker as part of the ArcticNet research program (N = 22, 5 stations were sampled in two separate years). At each station, 12 L Teflon-lined Niskin bottles (General Oceanics) mounted on the ship's rosette system were used to obtain samples from various depths, generally corresponding to the surface (~2-4 m), the subsurface chlorophyll maximum (SCM, the depth of maximum chlorophyll *a* fluorescence and hence algal biomass) and the lower oxycline (a zone of high heterotrophic respiration and organic carbon remineralization) and the bottom of the water column (~5-10 m above the seafloor). Subsamples were collected from the Niskin bottles into pre-cleaned amber Boston round bottles (Environmental Sampling Supplies) using the "clean hands-dirty hands" Hg sampling protocol. All samples were processed and preserved immediately after collection for analysis at a later date. Methylated Hg (MeHg + Me₂Hg) samples were preserved by acidification using trace-metal grade concentrated H₂SO₄ (equivalent to 0.4% of sample volume), the resulting low pH leading to the quantitative conversion of Me₂Hg to MeHg [Black et al., 2009]. Total Hg samples were preserved with the addition of trace-metal grade concentrated HCl (0.2% of sample volume). MeHg samples were preserved by freezing. In 2006, Me₂Hg was collected using the purge and trap technique whereby seawater collected in a custommade 1 L glass vessel was purged with ultra-high purity (UHP) N₂ gas and the Me₂Hg thus displaced from solution was collected in traps consisting of glass columns packed with Carbo trap. For the 2007 and 2010 data, Me₂Hg was calculated from the difference between methylated Hg and MeHg. Particles (seston) were collected in 2010 by filtering 5-9 L of seawater through 0.2 mm cellulose nitrate membranes. Filter membranes were frozen immediately after sample collection and freeze-dried once back on shore. Sample analysis for methylated Hg, MeHg and total Hg was carried out as described in Lehnherr et al. [2011]. MeHg was extracted from particles, by distilling the complete membrane filter, with Me²⁰¹Hg added as a species-specific internal standard prior to distillation to correct for procedural recoveries, similar to the procedure described for sediments in Lehnherr et al. [2012]. The distillate was then analyzed in the same fashion as with seawater samples.

Biota

МеНд	MeHg:THg	Reference
$(ng g^{-1} dw)$	(%)	
Mixed zooplankton		
4 ± 3^{A}	70 ± 38	[Campbell et al., 2005]
10±1	28	[Loseto et al., 2008]
95±19	75	[Loseto et al., 2008]
28 ± 0.8^{A}	100	[Campbell et al., 2005]
Copepods		
7±1	28	[Loseto et al., 2008]
11±1	34	[Loseto et al., 2008]
7±1	51	[<i>Pucko et al.</i> , 2014]
40±9	84	[<i>Pucko et al.</i> , 2014]
5-28	10-60	[Stern and Macdonald, 2005]
<1.5 ^A		[Jaeger et al., 2009]
2 ± 1^{A}	8	[Campbell et al., 2005]
Themisto libellula		
66±2	75	[Loseto et al., 2008]
<1.5 ^A		[<i>Jaeger et al.</i> , 2009]
Themisto abyssorum		
88±10	58	[<i>Pucko et al.</i> , 2014]
Chaethognats		_
17±4	74	[Pucko et al., 2014]

Table S2. Arctic zooplankton MeHg concentrations

^A ng g⁻¹ wet weight

Figure S4. Reservoirs of MeHg in plankton and bacteria (note log scale). Note that in the Arctic phytoplankton and zooplankton often has an inverted biomass pyramid and that the zooplankton biomass therefore is higher than that of phytoplankton [*ACIA*, 2005].



Model parameter description

Deposition $M_{dep-HgII} (Mg a^{-1})$	Hg ^{II} deposition to open ocean	30	[Dastoor and Durnford, 2014; Fisher et al., 2012]
$M_{dep-MeHg} (Mg \ a^{-1})$	MeHg deposition to open ocean based on fast redeposition of evaded Me ₂ Hg	$M_{eva-Me2Hg} imes F_{open-ocean-summer} imes F_{evasion-summer}$	(F _{open-ocean-summer} ; F _{evasion-summer} : Table 3)
$Meltwater \ M_{melt-HgII} \ (Mg \ a^{-1}) \ M_{melt-MeHg} \ (Mg \ a^{-1})$	Hg _{II} from meltwater MeHg from meltwater	$C_{melt-HgII} imes Prec imes (1-F_{open-ocean})$ $C_{melt-MeHg} imes Prec imes (1-F_{open-ocean})$	(F _{open-ocean} ; Prec: Table 3, C _{melt-HgII} : Table 4) (F _{open-ocean} ; Prec: Table 3, C _{melt-MeHg} : Table 4)
Multi-year sea-ice loss	П		
$M_{ice-HgII}(Mgy^{-1})$	Hg ⁿ from multi-year sea-ice loss	0.42	[Beattie et al., 2014]
$M_{ice-MeHg}(Mgy^{-1})$	MeHg from multi-year sea-ice loss	0.042	[Beattie et al., 2014]

 Table S3. Atmospheric deposition and melt water flux from snow and ice

Table S4. River and erosion fluxes

$M_{river-HgII} (Mg a^{-1})$	River Hg ^{II} discharge	50	[Dastoor and Durnford, 2014; Zhang et al., 2015]
$M_{river-MeHg} (Mg a^{-1})$	River MeHg discharge	$M_{river-HgII} imes F_{river-MeHg}$	
$M_{erosion-MeHg}$ (Mg a^{-1})	Erosion MeHg input	$M_{erosion-HgII} imes F_{MeHg-sediment}$	
$M_{erosion-HgII} (Mg a^{-1})$	Erosion Hg^{II} input	$TSS_{erosion} \times C_{erosion-HgII} \times 10^{-12}$	
$F_{river-MeHg}$	MeHg:Hg ^{II} fraction in river discharge	0.05 (0.01-0.10)	(Table 4)
$TSS_{erosion}(g a^{-1})$	TSS from erosion	430×10^{12}	(Figure 1)
$C_{erosion-HgII} (ng g^{-1})$	[Hg ^{II}] in eroded TSS	81 (61-114)	[<i>Leitch</i> , 2006]
F _{MeHg-sediment}	Fraction of eroded Hg ^{II} found as MeHg	0.01	[Hollweg et al., 2010]

M_{ox} (Mg a^{-1})	Oxidation flux	$M_{w-Hg0} \times (k_{phot-ox} + k_{dark-ox}) \times 365$	
M_{red} (Mg a^{-1})	Reduction flux	$M_{w-HgII} \times F_{red} \times (k_{bio-red} + k_{phot-red}) \times 3$	65
M_{met} (Mg a^{-1})	Methylation flux	$M_{w-HgII} \times k_{met} \times 365$	
$M_{dem} (Mg a^{-1})$	Demethylation flux	$M_{w-MeHg} \times (k_{phot-dem} + (k_{bio-dem} \times F_{rad-area})$	$()) \times 365$
M_{met2} (Mg a^{-1})	MeHg to Me ₂ Hg flux	$M_{w-MeHg} \times k_{met2} \times 365$	
M_{2dec} (Mg a^{-1})	Me ₂ Hg decomposition flux	$M_{w-Me2Hg} \times (k_{2dec} + k_{phot-2dec}) \times 365$	
M_w (Mg)	Mass of Hg species in specified reservoir		
$k_{phot-ox} (d^{-1})$	Photo-oxidation rate constant	$0.57 \times \text{RAD} \times (\text{F}_{\text{rad-area}})$	
$k_{phot-red} (d^{-1})$	Photo-reduction rate constant	$0.16 \times \text{RAD} \times (F_{\text{rad-area}})$	
$k_{dark-ox} (d^{-1})$	Dark oxidation rate constant	0.0086	[Soerensen et al., 2010]
$k_{bio-red} (d^{-1})$	Biotic reduction rate constant	$0.0450 \times \text{NPP}$	[Soerensen et al., 2010] (NPP: Table 3)
$k_{met} (d^{-1})$	Methylation rate constant	$0.0134 \times \text{NPP} + k_{\text{met-base}}$	[Lehnherr et al., 2011] (NPP: Table 3)
$k_{met-base} (d^{-1})$	Methylation base rate	0.0056	[Monperrus et al., 2007]
$k_{phot-dem} (m^2 E^{-1} d^{-1})$	Photo-demethylation rate constant	$PAR \times 3 \times 10^{-3} \times F_{summer}$	(see Table RATES)
$k_{bio-dem} (d^{-l})$	Biotic demethylation rate constant	0.07-0.18	
$k_{bio \ dem-base} \ (d^{-1})$	Demethylation base rate	0.05	[Monperrus et al., 2007]
$k_{met2} (d^{-1})$	MeHg to Me ₂ Hg rate constant	0.0016	(see Table RATES)
$k_{2dec} (d^{-1})$	Me ₂ Hg dark decomposition rate constant	0.0002	(see Table RATES)
$k_{phot-2dec} (m^2 E^{-1} d^{-1})$	Me ₂ Hg photodecomposition rate constant	$PAR \times 5 \times 10^{-4} \times F_{summer}$	50% of detection limit from <i>Black et al.</i> [2009]
F_{red}	Fraction of reducible Hg ^{II}	0.4	[Guentzel et al., 1996]
Frad-area	Fraction of ocean where light can penetrate	$F_{open-ocean} + F_{meltp}$	(F _{open-ocean} : Table 3)
	during summer (Apr-Sep) months		
F _{meltp}	Area of ocean with summer melt ponds	0.1	[Boetius et al., 2013]
F _{summer} /F _{winter}	Fraction of the year considered summer/winter	0.5	
$PAR(E m^{-2})$	Shortwave intensity as a function of time	$RAD \times 0.432$	
$RAD (W m^{-2})$	Shortwave intensity at specified depth intervals	$\frac{1}{1} \bullet \frac{R_i}{R_i} [e^{\eta x^1} - e^{-\eta x^2}]$	(RAD _i : Table 3)
		$x_2 - x_1 \eta$	
$x_1(m)$	Surface depth		
$x_2(m)$	Bottom depth		
$\eta (m^{-1})$	Extinction coefficient for radiation	$\eta_w + \eta_{Chl} C_{Chl} + \eta_{DOC} C_{DOC}$	
$\eta_w (m^{-1})$	Extinction coefficient for water	450 nm (vis) = 0.0145	
$\eta_{Chla} (m^{-1})$	Extinction coefficient for pigments	450 nm (vis) = 31	
$C_{Chla} (mg L^{-1})$	Average concentration of Chl a in mixed layer	0.6×10^{-3}	[Galand et al., 2008; Jin et al., 2012]
η_{DOC} (mg L^{-1})	Extinction coefficient for DOC	450 nm (vis) = 0.654	/ _
C_{DOC} (mg L^{-1})	Concentration of DOC in water column	0.8	[Dittmar and Kattner, 2003; Hansell et al., 2012]

 Table S6. Chemical transformation in sediment

$M_{sed-met} (Mg a^{-1})$	Sediment methylation flux	$C_{psed-HgII} \times k_{sed-met} \times 365 \ (d \ y^{-1})$	
$M_{sed-dem} (Mg a^{-1})$	Sediment demethylation flux	$C_{psed-MeHg} \times k_{sed-dem} \times 365 \ (d \ y^{-1})$	
$C_{psed-HgII}(pM)$	Porewater Hg ^{II} concentration	74.0 (14-258)	$C_{sed-HgII} \times 10^3 / kd_{HgII}$ (C _{sed} : Table 2)
$C_{psed-MeHg}(pM)$	Porewater MeHg concentration	2.4 (0.3-9.9)	$C_{sed-MeHg} \times 10^3 / kd_{MeHg}$ (C _{sed} : Table 2)
<i>kd</i> _{<i>HgII</i>}	Hg ^{II} partition coefficient	$10^{4.0} (10^{3.1} - 10^{4.2})$	[Hollweg et al., 2010; Schartup et al.,
_			2015; Sunderland et al., 2010]
kd_{MeHg}	MeHg partition coefficient	$10^{2.7} (10^{2.2} - 10^{3.1})$	[Hollweg et al., 2010; Schartup et al.,
0			2015; Sunderland et al., 2010]
$k_{sed-met} (d^{-1})$	Sediment methylation rate constant	0.03	[Hammerschmidt and Fitzgerald, 2008;
	-		Heyes et al., 2006; Hollweg et al., 2010]
$k_{sed-dem}(d^{-1})$	Sediment demethylation rate constant	0.58	Derived based on the sediment
. ,	-		MeHg:Hg ^{II} ratio

Table S7. Sediment burial

	Burial flux of Hg ^{II} and MeHg Hg species conc. in sediments Concentration of solids in benthic	$M_{sed} imes Burial / D_{sed}$ $V_{sed} imes Sedsolid imes C_{sed} imes 10^{-15} (Mg ng^{-1})$ (1-POR) imes DSS	Burial rate found in Table 3 1 × 10 ⁶ (g kg ⁻¹ m3 l ⁻¹) (C _{sed} : Table 2)
DSS (kg l^{1}) V_{sed} (m^{3}) $D_{sed-deep}$ (m) $D_{sed-shelf}$ (m)	sediments Density of sediment solids Sediment volume Depth of active deep ocean sediment Depth of active shelf sediment	1.5 $A_{sed} \times D_{sed}$ 0.01 0.02	(A _{sed} : Table 3) [Clough et al., 1997] [Clough et al., 1997; Kuzyk et al., 2013; Trefry et al., 2014]

Table S8. Diffusive transfer

Sediment-water int	terphase		
$ M_{sdiff-HgX} (Mg y^{-1}) F_{sw-diff} (ng cm^{-2} s^{-1}) $	Sediment-water net diffusion of Hg Diffusive flux based on Fick's law	$\frac{F_{sw-diff} \times A_{sed} \times 10^4 cm^2 m^{-2} \times (60 \times 60 \times 24 \times 36)}{POR \times D_{sw}} \times \frac{(C_{psed-Hg} - C_{w-Hg}) \times 1000}{D_{sed}}$	The diffusivity gradient is assumed to be in the top 1 cm of the sediment [Sunderland et al. 2010]
TOR	Tortuosity of sediments	$1 - \ln(POR^2)$	[<i>Gill et al.</i> , 1999]
POR	Porosity of sediments	0.68±0.10	[Hamden et al., 2013; Kuzyk et al., 2013]
$D_{sed} (cm^2 s^{-1})$	Sediment-water diffusion coefficient at temperature Tsed	$D_{sw25} / (1 + 0.048 \times (25 - T_{sed}))$	[Gill et al., 1999]
$D_{sw25} (cm^2 s^{-1})$	Sediment-water diffusion coefficient at 25 °C	$6 \times 10^{-6} (2 \times 10^{-6} - 12 \times 10^{-6})$	[Hollweg et al., 2010]
T_{sed} (°C)	Temperature of sediments	1	Approximation, a 5 °C change in average temperature results in 12-15% change in diffusion flux
A_{sed}	Sediment area		Table 3
Water column diffi	ision		
$M_{wdif-HgX} (Mg y^{-l})$	In water column eddy diffusion of X species $(Hg^{II}, Hg^{0}, MeHg, and Me_{2}Hg)$	$D_{w} \times \left(\frac{C_{w-HgX}}{DiffDepth_{z2} - DiffDepth_{z1}}\right) \times 60 \times 60 \times 24 \times 365$	
$D_w (m^2 s^{-1})$	Eddy diffusion coefficient	$0.8 \times 10^{-4} (< 0.05 \times 10^{-4} - 2.5 \times 10^{-4})$	[Padman and Dillon, 1991; Shaw and Stanton, 2014; Wallace et al., 1987]
DiffDepth _{so} (m)	Depth of diffusion gradient	20	
$DiffDepth_{ss}(m)$	Depth of diffusion gradient	140	
DiffDepth _{de} (m)	Depth of diffusion gradient	300	
z1 (m)	Upper diffusion depth		
z2 (m)	Lower diffusion depth		

Table S9. Gas exchange parameterization for Me₂Hg

$M_{eva-Me2Hg} (Mg y^{-1})$	Total Me ₂ Hg evasion flux (A (areas); F: in Table 3)	$F_{v} \times \left(A_{shelf} + A_{deep}\right) \times \left(\frac{F_{open-sum} + F_{open-sum}}{2}\right)$	$\frac{win}{2}$ × 10 ⁻¹⁵ × 24 × 365
$F_v (ng m^{-2} h^{-1})$	Me ₂ Hg air-sea exchange flux	$K_w(C_w - C_a / H'_X(T))$	
$C_a (ng m^{-3})$	Concentration of Me ₂ Hg in air	0.0038 ± 0.0031	[<i>Baya et al.</i> , 2015]
$C_w (ng m^{-3})$	Concentrations of Me ₂ Hg in PML		Table 1
$H'_{Me2Hg}(T)$	Temperature dependent dimensionless Henry's law constant for Me ₂ Hg	$\ln H' = (\frac{-2512.43}{T} + 7.27)$	[Lindqvist and Rodhe, 1985]
$K_w (m hr^{-1})$	Water-side mass transfer coefficient	$A \times u_{10}^2 (Sc_{HgX} / Sc_{CO_2})^{-0.5}$	[Wanninkhof, 1992]
A (unitless)	Constant based on the Weibull distribution of wind speeds over oceans	0.25	[Nightingale et al., 2000]
$u_{10} (m \ s^{-1})$	Average wind speed at 10 m above sea surface	5.6 (4-6)	[Nummelin et al., 2015; Serreze and Barry, 2005]
Sc_{CO_2}	Schmidt number for CO ₂	$0.11T'^2 - 6.16T' + 644.7$	[Poissant et al., 2000]
$T'(^{\circ}C)$	Water temperature	0	Approximation, a 5 °C change in temperature only lead to 4% change in evasion
TK (K)	Water temperature	T'+273.15	
Sc_{Hg}	Schmidt number for Me ₂ Hg	v/D	
$v(cm^2 s^{-1})$	Kinematic viscosity	$N/\rho = 0.017e^{(-0.025T')}$	[Poissant et al., 2000]
$\rho (mg \ cm^{-3})$	Seawater density	1025	
$D(cm^2 s^{-1})$	Me2Hg diffusivity based on Wilke-Chang	$7.4 \times 10^{-8} (\phi_w M_w)^{1/2} T$	[Wilke and Chang, 1955]
		$\mathbf{N}V_B^{0.6}$	
N(cP)	Viscosity of water	See Soerensen et al. (2010)	[Soerensen et al., 2010]
M_w (g mol ⁻¹)	Molecular weight of water	18.0	
$V_B (cm^3 mol^{-1})$	Molal volume of Me ₂ Hg at its normal boiling temperature	72.11	
$\phi_{\scriptscriptstyle W}$	Solvent association factor	2.26	[Hayduk and Laudie, 1974]

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